BARRIER INTEGRITY TESTING WITH A CONDENSER-CHAMBER TEWL INSTRUMENT

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1. Introduction

The aim of this study was to assess the performance of a condenser-chamber TEWL instrument for barrier integrity testing. OECD Test Guideline 428 stipulates barrier integrity testing to be carried out[1]. TEWL, electrical resistance and tritiated water procedures are recognised for such tests[2].

In a comprehensive study using an open-chamber TewaMeter, Netzlaff et al[3] found TEWL measurements to be of limited use for barrier integrity testing, being able to detect only severe damage in the samples they examined. Particular problems they identified included topically adhering water and the permeation of condensed water via capillary action through deliberately made pinholes in artificial and biological membranes. Our study assesses the extent to which such problems are reduced when using a condenser-chamber TEWL instrument, which offers (a) rapid drying of topically adhering water in its controlled, low humidity microclimate and (b) higher sensitivity.

2. Materials and Methods

Measurements were performed on artificial membranes (Sil-Tec from Technical Products Inc, USA and Teflon from Saarland University, Germany) and bio-membranes (excised human epidermis and excised human stratum corneum (SC)). Before the measurements, epidermis and SC samples were hydrated between wet filter paper sheets for 30 min, blotted dry, mounted on the Franz cell and left to acclimatise for 15 min. The system was then coupled to an AquaFlux measurement head and flux density time-series curves measured. Membrane damage was simulated by puncturing samples with a fine pin to produce perforations of ~50-100μm diameter.

TEWL was measured at room temperature using an AquaFlux AF200 instrument with a 9mm PermeGear Franz Diffusion Cell (PermeGear Inc, USA). A push-fit coupling between the TEWL measurement head and the Franz cell donor chamber was developed to give a reproducible, vapour-tight seal without the need to touch the membrane under test (**Figure 1**).

Figure 1: AquaFlux- Franz Cell Coupling

3. Results

3.1 Experiments with Sil-Tec membranes

Initial experiments used Sil-Tec membranes whose well controlled properties could be relied upon to verify the measurements. The controlled low-humidity microclimate within an AquaFlux measurement chamber offers a distinct advantage over conventional TEWL instruments, because any topical water evaporates quickly during the measurements. The measured flux curves clearly show the drying progress and therefore give quality control information for the tests. These points are illustrated in **Figure 2**, where three different curves are shown.
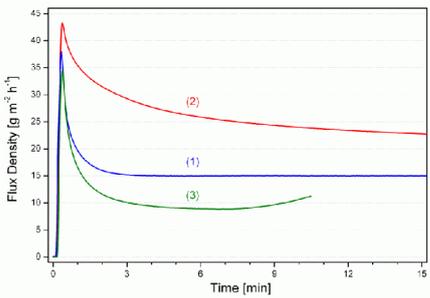

Figure 2: Franz-cell membrane tests using Sil-Tec membranes

Curve (1) shows rapid settling to a steady level. There was little donor-side moisture and the seal around the membrane was tight. **Curve (2)** settles more slowly as donor-side moisture evaporates. This causes the test to be prolonged, but the eventual result is valid. **Curve (3)** settles rapidly at first, then begins to rise again. This was found to be caused by a leaky seal around the membrane, resulting in a steadily increasing area of membrane contributing to the transport.

The validity of such measurements can be tested by correlating membrane diffusion resistance (ie 1/permeability) with membrane thickness, as illustrated in **Figure 3**.
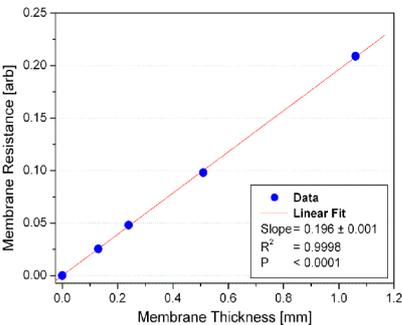

Figure 3: Diffusion Resistance Analysis for Sil-Tec membranes in the thickness range 0.13-1.06mm, showing an excellent linear correlation.

3.2 Experiments with Teflon Membranes

Similar measurements as published by Netzlaff el al[3] were performed on the same types of Teflon membranes, in order to assess condenser-chamber AquaFlux performance to quantify membrane damage. Netzlaff el al were able to detect single punctures of ~1mm diameter. Additional punctures were found to have little effect on the measured TEWL.

AquaFlux measurements were performed with a number of smaller punctures of 50-100μm diameter. The measured flux density was found to increase with membrane damage as measured by the number of membrane punctures, see **Figure 4**. The error bars relate to inconsistent puncture diameters rather than instrumental repeatability. Note that the sensitivity to membrane damage is highest at low damage.
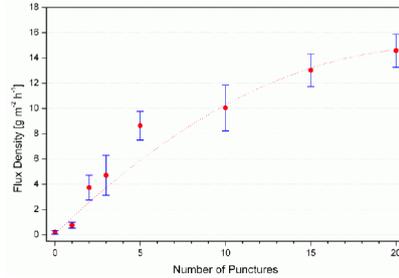

Figure 4: AquaFlux membrane damage measurements. Each point is the mean of 8 membranes tested. The error bars are ± 1 standard deviation and relate to inconsistent puncture diameters, not instrumental repeatability.

3.3 Experiments with SC

Measurements were performed on intact and deliberately damaged SC sheets from different donors. Typical flux density curves are presented in **Figure 5**.
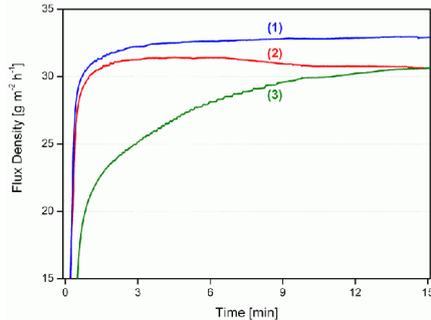

Figure 5: Flux density measurements on intact SC samples.

Curve (1) shows normal settling to a steady flux. **Curve (2)** shows the effect of donor-side moisture, which needs to evaporate before the flux settles to a steady level. This prolongs the test, but the result is valid. **Curve (3)** shows an anomalously slow rise to a steady flux, caused by poor contact between the receptor water and the lower surface of the membrane. The ability to inspect the flux time-series curves in this way is crucial for validating the tests.

Typical results of integrity tests on SC samples from different donors, before and after inflicting damage by means of a single puncture of 50-100μm diameter, are presented in **Figure 6**.
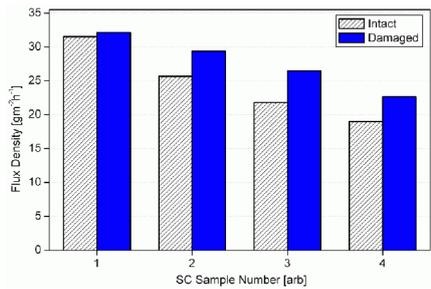

Figure 6: AquaFlux TEWL measurements on intact and damaged SC samples from different donors.

The effect of membrane damage is clearly visible in SC samples 2-4, where the intact membrane permeability is low. Sample 1 has a higher intact permeability and shows little effect from the additional damage.

3.4 Experiments with Epidermis

Similar experiments were performed with epidermis sheets. The results presented in **Figure 7** demonstrate the capability of the AquaFlux to differentiate between intact and damaged membranes.
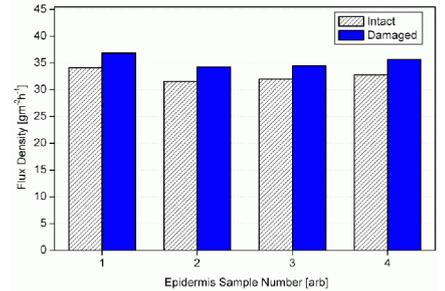

Figure 7: AquaFlux TEWL measurements on intact and damaged epidermis samples from different donors.

4. Repeatability of Measurements

Repeatability is an important instrumental attribute in membrane integrity testing, because it determines the extent to which small differences in readings are meaningful in terms of membrane permeability. The repeatability of the AquaFlux was assessed by performing multiple measurements on the same samples under otherwise similar conditions. Both SC and epidermis samples were used, with each sample tested 9 times. The Franz cell was uncoupled from the instrument for typically one minute between repeat measurements.
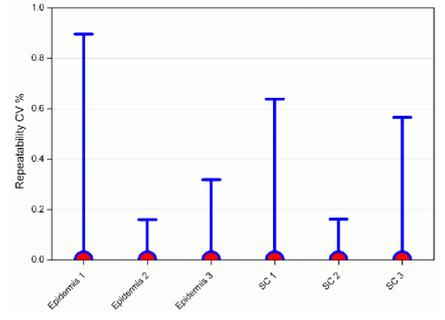

Figure 8: Coefficient of Variation CV% for nine repeat measurements on three epidermis and three SC samples.

The Coefficient of Variation (see **Figure 8**) was found to be less than 1% for all the samples tested. This corresponds to a standard deviation of less than 0.3 g m^{-2} h^{-1}. Therefore, the changes of flux density recorded in **Figures 6 and 7** are undoubtedly caused by sample damage and not by random fluctuations.

5. Summary

The main points arising from this study are:-

• A precise and leak-tight coupling between the Franz cell donor compartment and the TEWL measurement head is essential for repeatability.

• Experiments with Sil-Tec membranes of known thickness show that AquaFlux measurements correlate linearly with membrane diffusion resistance (ie 1/permeability), with a correlation coefficient close to unity (P<0.0001).

• Experiments with Teflon membranes show that AquaFlux measurements can detect membrane damage, with highest sensitivity for samples of lowest permeability. These findings are confirmed by the measurements on epidermis and SC.

• Measurements on SC and epidermis show that AquaFlux measurements are repeatable to better than 1% Coefficient of Variation.

6. Conclusions

In practical membrane integrity testing, the condenser-chamber method offers distinct advantages over other methods, as follows:-

1. The controlled condenser-chamber microclimate produces consistent measurement conditions irrespective of ambient humidity.

2. The controlled microclimate is also responsible for the outstanding repeatability of the tests.

3. The low humidity within the condenser-chamber causes topically adhering water to dry off quickly during measurements, thus reducing reliance on drying prior to measurement.

4. The recorded water vapour flux curves clearly show the drying progress and give quality control information for the tests.

5. The AquaFlux software can be set to terminate the test automatically when the quality criteria are met, thus ensuring that the tests are neither prematurely terminated nor are run for longer than necessary.

References

1. Skin absorption: In-vitro method. OECD Test Guideline 428, 2004.

2. Guidance document for the conduct of skin absorption studies. In: OECD Series on Testing and Assessment, No. 28, 2004.

3. F Netzlaff, KH Kostka, CM Lehr and UF Schaefer: TEWL measurements as a routine method for evaluating the integrity of epidermis sheets in static Franz type diffusion cells in vitro. Limitations shown by transport data testing. European Journal of Pharmaceutics and Biopharmaceutics, 63, 44-50, 2006.